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Cationic host defense (antimicrobial) peptides

Kelly L Brown and Robert EW Hancock

Members of the cationic host defense (antimicrobial) peptide family are widely distributed in nature, existing in organisms from insects to plants to mammals and non-mammalian vertebrates. Although many demonstrate direct antimicrobial activity against bacteria, fungi, eukaryotic parasites and/or viruses, it has been established that cationic peptides have a key modulatory role in the innate immune response. More recent evidence suggests that host defense peptides are effective adjuvants, are synergistic with other immune effectors, polarize the adaptive response, and support wound healing. In addition, the mechanisms of action are being unraveled, which support more effective implementation of derivatives of these endogenous peptides as therapeutic agents.

Addresses

Centre for Microbial Diseases and Immunity Research, University of British Columbia, Vancouver, British Columbia, V6T 1Z4, Canada

Corresponding author: Hancock, Robert EW (bob@cmdr.ubc.ca)

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Introduction: cationic peptides in host defense

Cationic amphipathic peptides are found in every complex species [1]. They are generally defined as having 12 to about 50 amino acids with 2–9 positively charged lysine or arginine residues and up to 50% hydrophobic amino acids. They fold into a variety of secondary structures (often after they insert into membrane bilayers) in which the charged and polar, and hydrophobic residues form patches on the surface of the molecule [2]. Early work with insects, amphibians and mammalian phagocytes demonstrated that they had direct antimicrobial activity against diverse microbes. More recently, it has become evident that they have a diverse range of functions in modulating immunity (Figure 1) which have an impact on infections and inflammation [1,3–5]. Therefore, although they are often termed antimicrobial peptides, we prefer the term used in this review to describe the breadth of their activities, namely cationic host defense peptides.

Peptides are expressed with elements of immunity

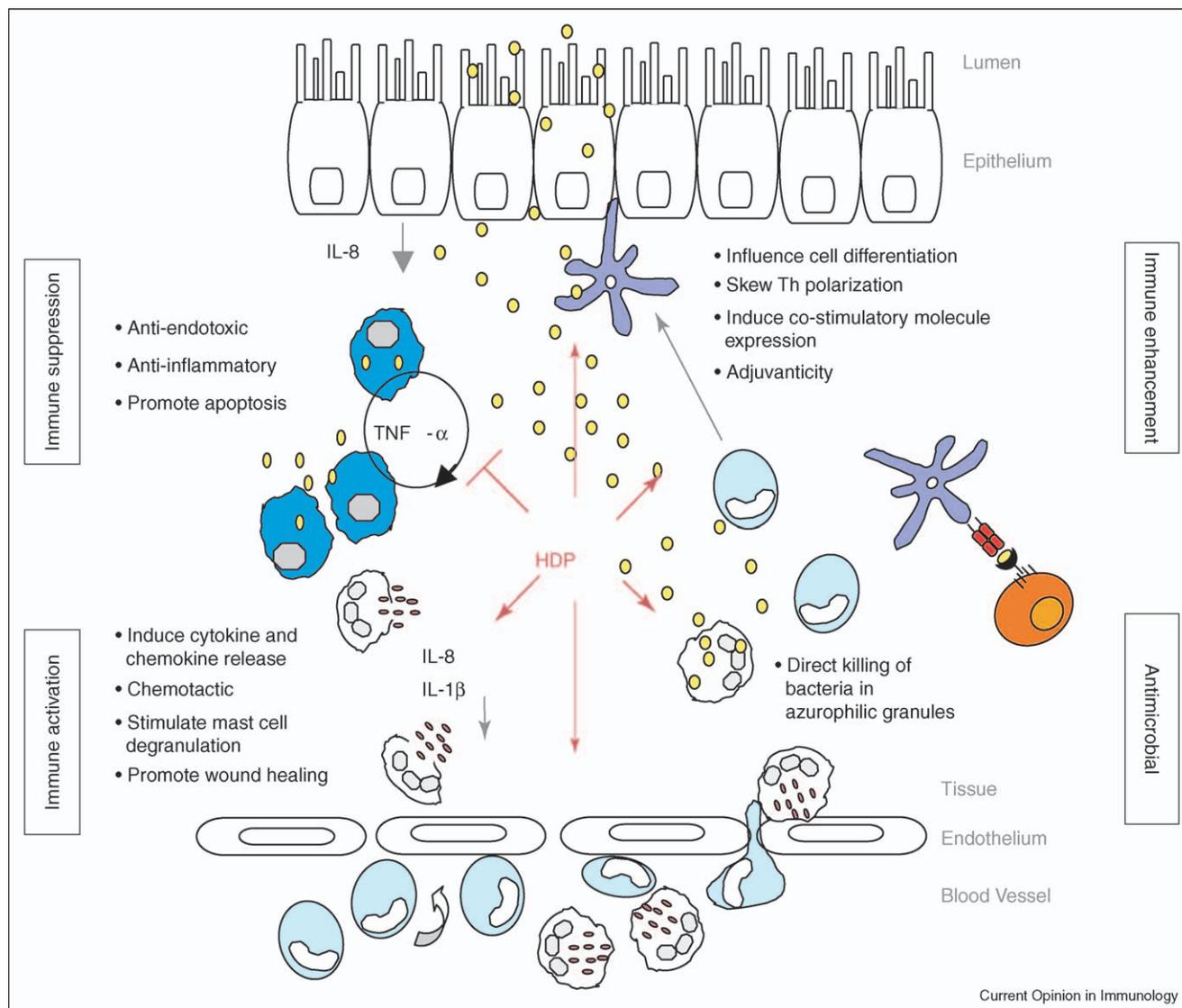
Innate immunity is the most ancient form of immune defense, conserved throughout the animal kingdom and vital to invertebrate host defenses. More recently discovered aspects of innate immunity — most notably, the family of Toll-like receptors (TLRs) — have illuminated the previously unrecognized complexity and heterogeneity within the innate immune system. Many factors support important and diverse roles for antimicrobial peptides in immunity: the robust membership, broad diversity in sequence and structure, thematic similarity of vertebrate and invertebrate antimicrobial peptides, wide distribution in cells of the immune system (leukocytes, Paneth cells) and in tissues that encounter bacterial infections (gut, trachea) and the observation that they might be either constitutive or demonstrate TLR- or inflammation-induced expression or secretion.

The expression of mature (biologically active) peptides requires proteolytic cleavage [6], might be constitutive or inducible and depends on species, tissue type, cellular lineages and/or differentiation state of the cell [7,8]. Gene expression and/or protein secretion are induced by factors such as bacterial products, injury and/or inflammatory stimuli. For example, hBD expression is upregulated in monocytes (hBD-1 and -2) exposed to bacteria, lipopolysaccharide (LPS) or IFN- γ [9,10], in keratinocytes (hBD-2–4) stimulated with TNF- α , IL-1 β , bacteria or IL-22 (for hBD-2 and -3) [11–13], and in intestinal, uterine or airway epithelial cells (hBD-1–3) stimulated with TLR agonists [14–17]. The 5' flanking sequences upstream of the cathelicidin coding sequence have several potential consensus sequences for transcription factors, including nuclear factor (NF) κ B, NF-IL6, acute phase response factor and IFN- γ response elements [18–20]. Synthesis of hBD-2 is induced in monocytes by IL-1 β and in intestinal epithelial cells by LPS or peptidoglycan and is dependent on NF κ B [14,21–23], whereas IL-22-induced hBD-2 and -3 expression in keratinocytes is dependent on the transcription factor STAT3 [13]. IL-1, TNF- α and TLR agonists also activate NF- κ B, the transcription factor that is responsible for the transcription of multiple inflammatory and immunity genes in mammals. These data demonstrate that transcriptional regulation of antimicrobial peptides is dependent on the stimulus and cell type and is regulated and/or coordinated with the expression of other entities of innate immunity and acute inflammation [19,20,24].

Peptides combat infection

The role for antimicrobial peptides in immunity is supported by *in vivo* evidence in humans and mice correlat-

Figure 1



Functions of cationic host defense peptides (HDPs). Within cytoplasmic granules, HDPs are antimicrobial agents that directly kill microbes, whereas secreted HDPs interact with cells of the innate immune system to indirectly eradicate pathogens (yellow circles represent bacteria/bacterial products). HDPs promote immune responses to infection yet simultaneously limit the magnitude of the inflammatory response (suppress production of pro-inflammatory cytokines such as $\text{TNF-}\alpha$). The multifunctional properties of HDPs make them attractive as therapeutic agents.

ing the expression of antimicrobial peptides with susceptibility or resistance to bacterial infections. Patients with specific granule deficiency syndrome lack α -defensins and suffer from severe and frequent bacterial infections [25]. Others with a condition known as morbus Kostmann suffer from frequent oral bacterial infections and severe periodontal disease which correlates with a deficiency in the human cathelicidin peptide LL-37 and human α -defensins (HNP1-3) [26]. Low expression of LL-37, human β -defensin (hBD)-2 and hBD-3 in skin lesions caused by atopic dermatitis coincides with enhanced susceptibility to skin infections [27,28]. Conversely, hBD-2 and hBD-3 expression are enhanced in psoriasis

[29] and in bronchoalveolar inflammation [30,31]. The expression of cathelicidins (LL-37 and mouse cathelicidin-related antimicrobial peptide [CRAMP]) in skin keratinocytes varies with infection and/or injury [32].

In rodent models, cathelicidins can control bacterial load and prevent mortality when administered after bacterial challenge [33–35]. Matrix metalloproteinase-7, β -defensin-1 and CRAMP gene knockout mice are more susceptible to, and fail to clear, infections [36–40]. It should be noted that, although these observations are consistent with a role for cationic peptides in host defense, they are not definitive in distinguishing between a direct

antimicrobial function for these peptides, an immunomodulatory role or both in contributing to defense. Further, these observations correlate the expression of cathelicidins and defensins with states of infection and inflammation and clearly establish a role for antimicrobial peptides in innate defense, clearance, prevention and protection against bacterial assault.

Direct antimicrobial activity

The protective effects of the peptides have been attributed, in part, to the direct antimicrobial killing properties of purified peptides against bacterial, fungal or viral pathogens observed *in vitro* [1,5,41]. Most active antimicrobial peptides are able to interact with bacterial membranes, as described by four separate models [2,5]. With substantial local perturbation of the cytoplasmic membrane bilayer, ion-permeable channels are created, leading to increased cytoplasmic membrane permeability and bacterial cell death. Conversely, a substantial number of antimicrobial peptides, including polyphemusin, a very potent horseshoe crab antimicrobial peptide [42], can traverse the membrane [2] and induce killing by acting on one or more anionic intracellular targets [42–45]. Although there is a broad range of potencies, virtually all cathelicidins and defensins have direct antimicrobial activity *in vitro* under the appropriate conditions. Although some peptides might retain bactericidal activity *in vitro* under physiological conditions, the direct killing activity is often antagonized by physiological salt conditions, monovalent and divalent ions and serum [46**]. There is no doubt that some peptides (e.g. α -defensins in neutrophil granules) are present at concentrations that virtually guarantee that they act in a directly microbicidal fashion; however, the low concentrations and antagonism by physiological salt concentrations (e.g. at mucosal surfaces) is consistent with an interpretation that at least some cationic peptides act by alternative means *in vivo*.

Immune system modifiers

More recent findings have established that, at physiological concentrations of peptides, salt and serum, antimicrobial peptides stimulate a broad range of biological effects relevant to inflammation, innate immunity and adaptive immunity [4,46**,47,48,49**] in innate immune cells (neutrophils and epithelial cells) and in cells that bridge the innate and adaptive immune systems (monocytes, macrophages and dendritic cells [DCs]). Mammalian host defense peptides have been shown to boost, inhibit or complement cellular functions such as chemotaxis, apoptosis, gene transcription and cytokine production [47,50]. Such biological effects probably promote bacterial clearance, although not via direct killing. Further, the evidence suggests that the peptides have roles in immunity beyond antimicrobial activity, in suppressing bacterial-induced cytokine production (anti-inflammatory), and stimulating wound healing, angiogenesis and adjuvanticity (all of which have been demon-

strated in animal or tissue models [51]), explaining the increasing use of the term ‘host defense peptides’.

Immune activation

Cathelicidins and defensins secreted at sites of infection and/or injury are chemotactic for effector cells, induce the transcription and secretion of chemokines and induce histamine release from mast cells [52–54]. Together, this promotes recruitment of innate and adaptive immune cells required for the cellular and humoral responses to pathogens. α -Defensins and hBD-2, -3 and -4 are chemotactic for monocytes, (memory and naïve) T cells and immature DCs [55–58]. Bovine, human, mouse and pig cathelicidins are chemotactic for virtually all subsets of peripheral blood cells *in vitro* [51,59] and *in vivo* [59,60]. For example, LL-37 induces IL-8 release which in turn promotes the chemotaxis of neutrophils and release of high concentrations of LL-37. A similar scenario is observed in invertebrates, where the circulating hemocytes migrate through chemotaxis to the site of injury, where they release peptides. Host defense peptides also stimulate the release of particular cytokines; for example, a peptide derived from the *Limulus polyphemus* anti-LPS factor induced the release of antiviral and immunomodulating cytokines, IFN- α , IFN- γ , IL-2 and IL-13, but not TNF- α or IL-6 [61], from human peripheral blood mononuclear cells. Further, this peptide increased survival in mice following a lethal dose of *Pseudomonas aeruginosa* which was correlated with diminished systemic TNF- α and elevated mRNA synthesis of IL-2, IL-12 and IL-13, but not IL-4 and IL-10, in the spleen and liver [61,62]. LL-37 induces the release of IL-6, IL-8, TNF- α , granulocyte-macrophage colony-stimulating factor and IL-1 β in human keratinocytes [63**] and enhances TNF- α and IL-6 secretion in LL-37-derived immature DCs [64]. Mouse β -defensin-2-matured DCs also secrete proinflammatory (Th1-polarizing) cytokines IL-12, IL-1 α , IL-1 β and IL-6 [65].

Inflammatory and immune suppression

Although mammalian host defense peptides directly stimulate certain innate immune functions considered to be proinflammatory (such as chemotaxis of leukocytes, and induction of cytokine, chemokine and histamine release), the peptides can also protect the host against detrimental, potentially lethal effects, particularly those resulting from an excessive TLR-induced inflammatory response [33,66**]. Cathelicidins suppress the transcription of the genes for proinflammatory cytokines (e.g. TNF- α and IL-6) and the release of proinflammatory mediators induced by LPS and other bacterial products, and prevent sepsis in rodents after bacterial challenge. The endotoxin-neutralizing activities of host defense peptides might also implicate cathelicidins in maintaining homeostasis, particularly in commensal-rich regions of the gut, and/or in dampening excessive inflammation. In addition, cathelicidins prevent the release of toxic components that

cause excess tissue damage and inflammation (e.g. the proline-arginine [PR]-rich porcine cathelicidin, PR-39, inhibits the production of reactive oxygen species, whereas bovine myeloid antimicrobial peptide-28 induces apoptosis of activated [infected] lymphocytes), and actively promote tissue regeneration. Cathelicidins and defensins promote cell proliferation, vasculogenesis and wound repair [67–70]. These neutralizing and resolving effects of peptides defend the host against the destructive components of inflammation.

Immune enhancement

There is also some indication that cathelicidins and defensins can act at the interface of innate and adaptive immunity [4], modulating DC function [64] and antigen-specific immune responses. LL-37 induces differentiation of primary monocyte-derived DCs, increases endocytic capacity, modifies phagocytic receptor expression and function, upregulates co-stimulatory molecule expression (CD86) and enhances Th1 cytokine secretion (IL-12) by LPS-stimulated DCs. Likewise, mouse BD2 stimulates DC maturation and upregulates the expression of co-stimulatory molecules (CD40, CD80 and CD86), major histocompatibility complex class II and chemokine receptor CCR7 (CCR7 can regulate trafficking towards T cell-rich areas) on DCs [65]. In this manner, the peptides might enhance aspects of adaptive immunity, supporting the differentiation of certain cell lineages and possibly altering the cytokine milieu and, in turn, the polarization (Th1 or Th2) of the response. IL-22 upregulates hBD-2 and -3 expression in keratinocytes [13], IL-4 release from Th2 cells and upregulation of major histocompatibility complex I antigen expression and acute phase proteins. The low expression of hBD-2 and -3 mRNA in skin lesions from patients with atopic dermatitis has been correlated with the elevated expression of Th2 cytokines [28]. These data provide evidence that host defense peptide expression, and potentially function, correlates with aspects of the adaptive immune response. Further, LL-37 has synergy with cytokines, such as granulocyte-macrophage colony-stimulating factor and IL-1 β [46,66**], thereby enabling low concentrations of the peptide to influence the immune response. Used as adjuvants, LL37, CRAMP and mouse BD2 enhanced antigen-specific humoral and cellular responses [59,71**,72]. These peptides might be effective adjuvants as a result of their ability to elicit several responses, including the recruitment, differentiation and activation of effector cells at the site of infection. However, the effects of peptides on lymphocyte function (B cell activation and antibody production, cytotoxic T cell and natural killer cell killing and Th cell function) are, as yet, poorly described in the literature.

Mechanism of immune modulation

The biological effects of host defense peptides, similarly to their expression and secretion, are often induced by inflammatory stimuli (including conserved microbial

components of endogenous or pathogenic origin) and are influenced by the physiological setting, including the concentration of the peptide, the cellular environment and soluble components of the extracellular milieu. The mechanism by which host defense peptides exert immune-modulating effects probably involves multiple mechanisms, including direct binding to endotoxic LPS and known or putative surface receptors and/or intracellular signaling molecules and receptors. It has been reported that cathelicidins bind to a variety of receptors [73–75,76*,77], activate components of the mitogen-activated protein kinase signal transduction pathways [59,75,77,78], induce Ca²⁺ mobilization [52,59], bind to SH3-domain-containing proteins [79,80] and inhibit LPS-induced NF κ B translocation [66**,81]. Mouse BD2 activates NF κ B and might function as an endogenous ligand of TLR4 signaling [65]. The ligand–receptor interactions of cathelicidins are not well understood, and, although some functions are dependent on known receptors (e.g. LL-37 directly mediates chemotaxis of human peripheral blood neutrophils, monocytes and T cells through formyl peptide receptor-like 1 [73]), other biological functions are not associated with known receptors.

Therapeutics

Functional similarities among the antimicrobial host defense peptides of distant evolutionary species indicate that the study of both vertebrate and invertebrate peptides could permit the development of new design templates for anti-infectious agents in humans [5,82]. Although not widely investigated at present, there are an increasing number of reports of the immunomodulatory effects of natural and synthetic host defense peptides on mammalian hosts, including neutralization of LPS, induction of signal transduction, gene transcription and release of reactive oxygen species [82,83], and interest is high for developing these as a novel therapy for human infectious diseases, through the selective boosting of innate immunity [51]. Several companies are actively pursuing the host defense peptides as novel antimicrobial therapeutics, with the indolicidin-like peptides of Migenix (formerly Micrologix) having advanced to Phase IIIb clinical trials for prevention of catheter-associated infections [84*]. The functional redundancy between species, the antiseptic and antimicrobial activities, adjuvant properties and low toxicity make them attractive therapeutic agents [1,48,82,84*]. Further, functionally active domains are apparently localized to different regions of the peptide (demonstrated for the antimicrobial and immunomodulatory functions), enabling the development of peptides with potent and specific functions [51,63**].

Conclusions

In summary, there is emerging evidence that host defense peptides actively participate in all stages of host immune defenses: exerting antimicrobial activity through direct killing and/or stimulation of biological functions in

immune effector cells during the inflammatory and immune response, by communication with cells at the interface of the innate and adaptive immune system, and in the control and resolution of the inflammatory response.

A more detailed analysis of host defense peptides will aid our understanding of how these peptides participate in the recognition and neutralization of pathogens, which will assist in the development of a new anti-infective therapeutic strategy. These studies will expand on current structure–function analyses and will build on reports of immune modulating functions by elucidating the mechanisms of action of the peptides. Of highest importance, future studies will exploit and build on the diverse nature of peptides and adhere to physiologically relevant conditions, ultimately validating, *in vivo*, host defense peptide functions that protect against bacterial challenge and suppress potentially harmful inflammation.

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References and recommended reading

Papers of particular interest, published within the period of review, have been highlighted as:

- of special interest
- of outstanding interest

1. Hancock REW, Diamond G: **The role of cationic antimicrobial peptides in innate host defences.** *Trends Microbiol* 2000, **8**:402-410.
2. Hancock REW, Rozek A: **Role of membranes in the activities of antimicrobial cationic peptides.** *FEMS Microbiol Lett* 2002, **206**:143-149.
3. Yang D, Biragyn A, Kwak LW, Oppenheim JJ: **Mammalian defensins in immunity: more than just microbicidal.** *Trends Immunol* 2002, **23**:291-296.
4. Yang D, Biragyn A, Hoover DM, Lubkowski J, Oppenheim JJ: **Multiple roles of antimicrobial defensins, cathelicidins and eosinophil-derived neurotoxin in host defense.** *Annu Rev Immunol* 2004, **22**:181-215.
5. Zasloff M: **Antimicrobial peptides of multicellular organisms.** *Nature* 2002, **415**:389-395.
6. Uzzell T, Stolzenberg ED, Shinnar AE, Zasloff M: **Hagfish intestinal antimicrobial peptides are ancient cathelicidins.** *Peptides* 2003, **24**:1655-1667.
7. Heilborn JD, Nilsson MF, Jimenez CI, Sandstedt B, Borregaard N, Tham E, Sorensen OE, Weber G, Stahle M: **Antimicrobial protein hCAP18/LL-37 is highly expressed in breast cancer and is a putative growth factor for epithelial cells.** *Int J Cancer* 2005, **114**:713-719.
8. Islam D, Bandholtz L, Nilsson J, Wigzell H, Christensson B, Agerberth B, Gudmundsson GH: **Downregulation of bactericidal peptides in enteric infections: a novel immune escape mechanism with bacterial DNA as a potential regulator.** *Nat Med* 2001, **7**:180-185.
9. Duits LA, Ravensbergen B, Rademaker M, Hiemstra PS, Nibbering PH: **Expression of β -defensin 1 and 2 mRNA by human monocytes, macrophages and dendritic cells.** *Immunology* 2002, **106**:517-525.
10. Fang XM, Shu Q, Chen QX, Book M, Sahl HG, Hoefl A, Stuber F: **Differential expression of α - and β -defensins in human peripheral blood.** *Eur J Clin Invest* 2003, **33**:82-87.
11. Harder J, Bartels J, Christophers E, Schroder JM: **Isolation and characterization of human β -defensin-3, a novel human inducible peptide antibiotic.** *J Biol Chem* 2001, **276**:5707-5713.
12. Harder J, Meyer-Hoffert U, Wehkamp K, Schwichtenberg L, Schroder JM: **Differential gene induction of human beta-defensins (hBD-1, -2, -3, and -4) in keratinocytes is inhibited by retinoic acid.** *J Invest Dermatol* 2004, **123**:522-529.
13. Wolk K, Kunz S, Witte E, Friedrich M, Asadullah K, Sabat R: **IL-22 increases the innate immunity of tissues.** *Immunity* 2004, **21**:241-254.
14. Vora P, Youdim A, Thomas LS, Fukata M, Tesfay SY, Lukasek K, Michelsen KS, Wada A, Hirayama T, Arditi M, Abreu MT: **Beta-defensin-2 expression is regulated by TLR signaling in intestinal epithelial cells.** *J Immunol* 2004, **173**:5398-5405.
15. Schaefer TM, Fahey JV, Wright JA, Wira CR: **Innate immunity in the human female reproductive tract: antiviral response of uterine epithelial cells to the TLR3 agonist poly(I:C).** *J Immunol* 2005, **174**:992-1002.
16. Proud D, Sanders SP, Wiehler S: **Human rhinovirus infection induces airway epithelial cell production of human beta-defensin 2 both *in vitro* and *in vivo*.** *J Immunol* 2004, **172**:4637-4645.
17. Platz J, Beisswenger C, Dalpke A, Koczulla R, Pinkenburg O, Vogelmeier C, Bals R: **Microbial DNA induces a host defense reaction of human respiratory epithelial cells.** *J Immunol* 2004, **173**:1219-1223.
18. Gudmundsson GH, Agerberth B, Odeberg J, Bergman T, Olsson B, Salcedo R: **The human gene FALL-39 and processing of the cathelin precursor to the antibacterial peptide LL-37 in granulocytes.** *Eur J Biochem* 1996, **238**:325-332.
19. Tomasinsig L, Zanetti M: **The cathelicidins – structure, function and evolution.** *Curr Protein Pept Sci* 2005, **6**:23-34.
20. Zhao C, Ganz T, Lehrer RI: **Structures of genes for two cathelin-associated antimicrobial peptides: prophenin-2 and PR-39.** *FEBS Lett* 1995, **376**:130-134.
21. Liu L, Roberts AA, Ganz T: **By IL-1 signaling, monocyte-derived cells dramatically enhance the epidermal antimicrobial response to lipopolysaccharide.** *J Immunol* 2003, **170**:575-580.
22. Hertz CJ, Wu Q, Porter EM, Zhang YJ, Weismuller KH, Godowski PJ, Ganz T, Randell SH, Modlin RL: **Activation of Toll-like receptor 2 on human tracheobronchial epithelial cells induces the antimicrobial peptide human defensin-2.** *J Immunol* 2003, **171**:6820-6826.
23. Wang X, Zhang Z, Louboutin JP, Moser C, Weiner DJ, Wilson JM: **Airway epithelia regulate expression of human beta-defensin 2 through Toll-like receptor 2.** *FASEB J* 2003, **17**:1727-1729.
24. Frohm Nilsson M, Sandstedt B, Sorensen O, Weber G, Borregaard N, Stahle-Bäckdahl M: **The human cationic antimicrobial protein (hCAP18), a peptide antibiotic, is widely expressed in human squamous epithelia and colocalizes with interleukin-6.** *Infect Immun* 1999, **67**:2561-2566.
25. Ganz T, Metcalf JA, Gallin JI, Boxer LA, Lehrer RI: **Microbicidal/cytotoxic proteins of neutrophils are deficient in two disorders: Chediak-Higashi syndrome and 'specific' granule deficiency.** *J Clin Invest* 1988, **82**:552-556.
26. Putsep K, Carlsson G, Boman HG, Andersson M: **Deficiency of antibacterial peptides in patients with morbus Kostmann: an observation study.** *Lancet* 2002, **360**:1144-1149.
27. Ong PY, Ohtake T, Brandt C, Strickland I, Boguniewicz M, Ganz T, Gallo RL, Leung DY: **Endogenous antimicrobial peptides and skin infections in atopic dermatitis.** *N Engl J Med* 2002, **347**:1151-1160.

28. Nomura I, Goleva E, Howell MD, Hamid QA, Ong PY, Hall CF, Darst MA, Gao B, Boguniewicz M, Travers JB, Leung DY: **Cytokine milieu of atopic dermatitis, as compared to psoriasis, skin prevents induction of innate immune response genes.** *J Immunol* 2003, **171**:3262-3269.
29. Harder J, Schroder JM: **Psoriatic scales: a promising source for the isolation of human skin-derived antimicrobial proteins.** *J Leukoc Biol* 2005, **77**:476-486.
30. Hiratsuka T, Mukae H, Iiboshi H, Ashitani J, Nabeshima K, Minematsu T, Chino N, Ihi T, Kohno S, Nakazato M: **Increased concentrations of human beta-defensins in plasma and bronchoalveolar lavage fluid of patients with diffuse panbronchiolitis.** *Thorax* 2003, **58**:425-430.
31. Ross DJ, Cole AM, Yoshioka D, Park AK, Belperio JA, Laks H, Strieter RM, Lynch JP III, Kubak B, Ardehali A, Ganz T: **Increased bronchoalveolar lavage human beta-defensin type 2 in bronchiolitis obliterans syndrome after lung transplantation.** *Transplantation* 2004, **78**:1222-1224.
32. Dorschner RA, Pestonjamas V, Tamakuwala S, Ohtake T, Rudisill J, Nizet V, Agerberth B, Gudmundsson GH, Gallo RL: **Cutaneous injury induces the release of cathelicidin anti-microbial peptides active against group A *Streptococcus*.** *J Invest Dermatol* 2001, **117**:91-97.
33. Scott MG, Davidson DJ, Gold MR, Bowdish D, Hancock REW: **The human antimicrobial peptide LL-37 is a multifunctional modulator of innate immune responses.** *J Immunol* 2002, **169**:3883-3891.
34. Fukumoto K, Nagaoka I, Yamataka A, Kobayashi H, Yanai T, Kato Y, Miyano T: **Effect of antibacterial cathelicidin peptide CAP18/LL-37 on sepsis in neonatal rats.** *Pediatr Surg Int* 2005, **21**:20-24.
35. Giacometti A, Cirioni O, Ghiselli R, Mocchegiani F, D'Amato F, Circo R, Orlando F, Skerlavaj B, Silvestri C, Saba V *et al.*: **Cathelicidin peptide sheep myeloid antimicrobial peptide-29 prevents endotoxin-induced mortality in rat models of septic shock.** *Am J Respir Crit Care Med* 2004, **169**:187-194.
36. Moser C, Weiner DJ, Lysenko E, Bals R, Weiser JN, Wilson JM: **β -Defensin 1 contributes to pulmonary innate immunity in mice.** *Infect Immun* 2002, **70**:3068-3072.
37. Rosenberger CM, Gallo RL, Finlay BB: **Interplay between antibacterial effectors: a macrophage antimicrobial peptide impairs intracellular *Salmonella* replication.** *Proc Natl Acad Sci USA* 2004, **101**:2422-2427.
38. Iimura M, Gallo RL, Hase K, Miyamoto Y, Eckmann L, Kagnoff MF: **Cathelicidin mediates innate intestinal defense against colonization with epithelial adherent bacterial pathogens.** *J Immunol* 2005, **174**:4901-4907.
39. Nizet V, Ohtake T, Lauth X, Trowbridge J, Rudisill J, Dorschner RA, Pestonjamas V, Piraino J, Huttner K, Gallo RL: **Innate antimicrobial peptide protects the skin from invasive bacterial infection.** *Nature* 2001, **414**:454-457.
40. Salzman NH, Ghosh D, Huttner KM, Paterson Y, Bevins CL: **Protection against enteric salmonellosis in transgenic mice expressing a human intestinal defensin.** *Nature* 2003, **422**:522-526.
41. Bader MW, Navarre WW, Shiau W, Nikaido H, Frye JG, McClelland M, Fang FC, Miller SI: **Regulation of *Salmonella typhimurium* virulence gene expression by cationic antimicrobial peptides.** *Mol Microbiol* 2003, **50**:219-230.
42. Zhang L, Scott MG, Yan H, Mayer LD, Hancock REW: **Interaction of polyphemusin I and structural analogs with bacterial membranes, lipopolysaccharide, and lipid monolayers.** *Biochemistry* 2000, **39**:14504-14514.
43. Kragol G, Lovas S, Varadi G, Condie BA, Hoffmann R, Otvos L Jr: **The antibacterial peptide pyrrolicin inhibits the ATPase actions of DnaK and prevents chaperone-assisted protein folding.** *Biochemistry* 2001, **40**:3016-3026.
44. Patrzykat A, Friedrich CL, Zhang L, Mendoza V, Hancock REW: **Sublethal concentrations of pleurocidin-derived antimicrobial peptides inhibit macromolecular synthesis in *Escherichia coli*.** *Antimicrob Agents Chemother* 2002, **46**:605-614.
45. Hong RW, Shchepetov M, Weiser JN, Axelsen PH: **Transcriptional profile of the *Escherichia coli* response to the antimicrobial insect peptide cecropin A.** *Antimicrob Agents Chemother* 2003, **47**:1-6.
46. Bowdish DM, Davidson DJ, Lau YE, Lee K, Scott MG, Hancock REW: **Impact of LL-37 on anti-infective immunity.** *J Leukoc Biol* 2005, **77**:451-459.
- The authors provide the first demonstration that a synthetic cationic host defense peptide lacking antimicrobial activity is protective in animal models of infection.
47. Finlay BB, Hancock RE: **Can innate immunity be enhanced to treat infections?** *Nat Rev Microbiol* 2004, **2**:497-504.
48. Hiemstra PS, Fernie-King BA, McMichael J, Lachmann PJ, Sallenave JM: **Antimicrobial peptides: mediators of innate immunity as templates for the development of novel anti-infective and immune therapeutics.** *Curr Pharm Des* 2004, **10**:2891-2905.
49. Selsted ME, Ouellette AJ: **Mammalian defensins in the antimicrobial immune response.** *Nat Immunol* 2005, **6**:551-557.
- This comprehensive and current review focuses on the roles of mammalian defensins in the immune response.
50. Salzet M: **Antimicrobial peptides are signaling molecules.** *Trends Immunol* 2002, **23**:283-284.
51. Bowdish DM, Davidson DJ, Scott MG, Hancock REW: **Immunomodulatory activities of small host defense peptides.** *Antimicrob Agents Chemother* 2005, **49**:1727-1732.
52. Niyonsaba F, Someya A, Hirata M, Ogawa H, Nagaoka I: **Evaluation of the effects of peptide antibiotics human beta-defensins-1/2 and LL-37 on histamine release and prostaglandin D(2) production from mast cells.** *Eur J Immunol* 2001, **31**:1066-1075.
53. Befus AD, Mowat C, Gilchrist M, Hu J, Solomon S, Bateman A: **Neutrophil defensins induce histamine secretion from mast cells: mechanisms of action.** *J Immunol* 1999, **163**:947-953.
54. Niyonsaba F, Iwabuchi K, Someya A, Hirata M, Matsuda H, Ogawa H, Nagaoka I: **A cathelicidin family of human antibacterial peptide LL-37 induces mast cell chemotaxis.** *Immunology* 2002, **106**:20-26.
55. Territo MC, Ganz T, Selsted ME, Lehrer RI: **Monocyte-chemotactic activity of defensins from human neutrophils.** *J Clin Invest* 1989, **84**:2017-2020.
56. Chertov O, Michiel DF, Xu L, Wang JM, Tani K, Murphy WJ, Longo DL, Taub DD, Oppenheim JJ: **Identification of defensin-1, defensin-2, and CAP37/azurocidin as T-cell chemoattractant proteins released from interleukin-8-stimulated neutrophils.** *J Biol Chem* 1996, **271**:2935-2940.
57. Yang D, Chen Q, Chertov O, Oppenheim JJ: **Human neutrophil defensins selectively chemoattract naive T and immature dendritic cells.** *J Leukoc Biol* 2000, **68**:9-14.
58. Garcia JR, Krause A, Schulz S, Rodriguez-Jimenez FJ, Kluver E, Adermann K, Forssmann U, Frimpong-Boateng A, Bals R, Forssmann WG: **Human beta-defensin 4: a novel inducible peptide with a specific salt-sensitive spectrum of antimicrobial activity.** *FASEB J* 2001, **15**:1819-1821.
59. Kurosaka K, Chen Q, Yarovinsky F, Oppenheim JJ, Yang D: **Mouse cathelin-related antimicrobial peptide chemoattracts leukocytes using formyl peptide receptor-like 1/mouse formyl peptide receptor-like 2 as the receptor and acts as an immune adjuvant.** *J Immunol* 2005, **174**:6257-6265.
60. Ohgami K, Iliava IB, Shiratori K, Isogai E, Yoshida K, Kotake S, Nishida T, Mizuki N, Ohno S: **Effect of human cationic antimicrobial protein 18 peptide on endotoxin-induced uveitis in rats.** *Invest Ophthalmol Vis Sci* 2003, **44**:4412-4418.
61. Vallespi MG, Glaria LA, Reyes O, Garray HE, Ferrero J, Araña MJ: **A Limulus antilipopolysaccharide factor-derived peptide exhibits a new immunological activity with potential applicability in infectious diseases.** *Clin Diagn Lab Immunol* 2000, **7**:669-675.

62. Vallespi MG, Alvarez-Obregon JC, Rodriguez-Alonso I, Montero T, Garay H, Reyes O, Arana MJ: **A Limulus anti-LPS factor-derived peptide modulates cytokine gene expression and promotes resolution of bacterial acute infection in mice.** *Int Immunopharmacol* 2003, **3**:247-256.
63. Braff M, Hawkins MA, Di Nardo A, Lopez-Garcia B, Howell MD, ●● Wong C, Lin K, Streib JE, Dorschner R, Leung DYM, Gallo RL: **Structure-function relationships among human cathelicidin peptides: dissociation of antimicrobial properties from host immunostimulatory activities.** *J Immunol* 2005, **174**:4271-4278.
- The authors demonstrate that immune modulating and antimicrobial activity are associated with different regions of the peptide, which has a significant impact on the design of immunomodulatory peptide therapeutics.
64. Davidson DJ, Currie AJ, Reid GS, Bowdish DM, MacDonald KL, Ma RC, Hancock REW, Speert DP: **The cationic antimicrobial peptide LL-37 modulates dendritic cell differentiation and dendritic cell-induced T cell polarization.** *J Immunol* 2004, **172**:1146-1156.
65. Biragyn A, Ruffini PA, Leifer CA, Klyushnenkova E, Shakhov A, Chertov O, Shirakawa AK, Farber JM, Segal DM, Oppenheim JJ, Kwak LW: **Toll-like receptor 4-dependent activation of dendritic cells by β -defensin 2.** *Science* 2002, **298**:1025-1029.
66. Mookherjee N, Brown KL, Bowdish DME, Doria S, Falsafi R, ●● Hokamp K, Roche FM, Mu R, Doho GH, Pistolic J *et al.*: **Modulation of the Toll-like receptor-mediated inflammatory response by the endogenous human host defence peptide LL-37.** *J Immunol* 2006, in press.
- This report demonstrates the anti-endotoxic effect of LL-37 and provides evidence that LL-37 selectively modulates LPS-induced transcription, in part, through inhibition of LPS-induced NF- κ B translocation.
67. Murphy CJ, Foster BA, Mannis MJ, Selsted ME, Reid TW: **Defensins are mitogenic for epithelial cells and fibroblasts.** *J Cell Physiol* 1993, **155**:408-413.
68. Aarbiou J, Ertmann M, van Wetering S, van Noort P, Rook D, Rabe KF, Litvinov SV, van Krieken JH, de Boer WI, Hiemstra PS: **Human neutrophil defensins induce lung epithelial cell proliferation *in vitro*.** *J Leukoc Biol* 2002, **72**:167-174.
69. Aarbiou J, Verhoosel RM, Van Wetering S, De Boer WI, Van Krieken JH, Litvinov SV, Rabe KF, Hiemstra PS: **Neutrophil defensins enhance lung epithelial wound closure and mucin gene expression *in vitro*.** *Am J Respir Cell Mol Biol* 2004, **30**:193-201.
70. Chavakis T, Cines DB, Rhee JS, Liang OD, Schubert U, Hammes HP, Higazi AA, Nawroth PP, Preissner KT, Bdeir K: **Regulation of neovascularization by human neutrophil peptides (alpha-defensins): a link between inflammation and angiogenesis.** *FASEB J* 2004, **18**:1306-1308.
71. An LL, Yang YH, Ma XT, Lin YM, Li G, Song YH, Wu KF: **LL-37 ●● enhances adaptive antitumor immune response in a murine model when genetically fused with M-CSFR (J6-1) DNA vaccine.** *Leuk Res* 2005, **29**:535-543.
- Although demonstrated previously for CRAMP and mBD-2 [59,72], this is the first report to show that the human cathelicidin LL-37 has adjuvant activity.
72. Biragyn A, Belyakov IM, Chow Y-H, Dimitrov DS, Berzofsky JA, Kwak LW: **DNA vaccines encoding human immunodeficiency virus-1 glycoprotein 120 fusions with proinflammatory chemoattractants induce systemic and mucosal immune responses.** *Blood* 2002, **100**:1153-1159.
73. Yang D, Chen Q, Schmidt AP, Anderson GM, Wang JM, Wooters J, Oppenheim JJ, Chertov O: **LL-37, the neutrophil granule- and epithelial cell-derived cathelicidin, utilizes formyl peptide receptor-like 1 (FPRL1) as a receptor to chemoattract human peripheral blood neutrophils, monocytes, and T cells.** *J Exp Med* 2000, **192**:1069-1074.
74. Niyonsaba F, Iwabuchi K, Matsuda H, Ogawa H, Nagaoka I: **Epithelial cell-derived human beta-defensin-2 acts as a chemotaxin for mast cells through a pertussis toxin-sensitive and phospholipase C-dependent pathway.** *Int Immunol* 2002, **14**:421-426.
75. Tjabringa GS, Aarbiou J, Ninaber DK, Drijfhout JW, Sorensen OE, Borregaard N, Rabe KF, Hiemstra PS: **The antimicrobial peptide LL-37 activates innate immunity at the airway epithelial surface by transactivation of the epidermal growth factor receptor.** *J Immunol* 2003, **171**:6690-6696.
76. Ellsner A, Duncan M, Gavrillin M, Wewers MD: **A novel P2X7 ● receptor activator, the human cathelicidin-derived peptide LL37, induces IL-1 β processing and release.** *J Immunol* 2004, **172**:4987-4994.
- The authors provide evidence that LL-37 transactivates a cell surface receptor (P2X7) to induce ATP release and the subsequent processing and release of IL-1 β .
77. Lau YE, Rozek A, Scott MG, Goosney DL, Davidson DJ, Hancock REW: **Interaction and cellular localization of the human host defense peptide LL-37 with lung epithelial cells.** *Infect Immun* 2005, **73**:583-591.
78. Bowdish D, Davidson DJ, Speert DP, Hancock REW: **The human cationic peptide LL-37 induces activation of the extracellular signal-regulated kinase and p38 kinase pathways in primary human monocytes.** *J Immunol* 2004, **172**:3758-3765.
79. Chan YR, Gallo RL: **PR-39, a syndecan-inducing antimicrobial peptide, binds and affects p130 (Cas).** *J Biol Chem* 1998, **273**:28978-28985.
80. Shi J, Ganz T: **The role of protegrins and other elastase-activated polypeptides in the bactericidal properties of porcine inflammatory fluids.** *Infect Immun* 1998, **66**:3611-3617.
81. Gao Y, Lecker S, Post MJ, Hietaranta AJ, Li J, Volk R, Li M, Sato K, Saluja AK, Steer ML *et al.*: **Inhibition of ubiquitin-proteasome pathway-mediated I κ B α degradation by a naturally occurring antibacterial peptide.** *J Clin Invest* 2000, **106**:439-448.
82. McPhee JB, Scott MG, Hancock REW: **Design of host defence peptides for antimicrobial and immunity enhancing activities.** *Comb Chem High Throughput Screen* 2005, **8**:257-272.
83. Hancock REW, Brown KL, Mookherjee N: **Host defence peptides from invertebrates — emerging antimicrobial strategies.** *Immunobiology* 2005, in press.
84. Zhang L, Falla TJ: **Cationic antimicrobial peptides — an update. ● Expert Opin Investig Drugs** 2004, **13**:97-106.
- This reviews antimicrobial peptides used in current drug design, and the commercial potential thereof.

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